

# Calcium Influx via TRP Channels Is Required to Maintain PIP<sub>2</sub> Levels in *Drosophila* Photoreceptors

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## Summary

The *trp* (*transient receptor potential*) gene encodes a Ca<sup>2+</sup> channel responsible for the major component of the phospholipase C (PLC) mediated light response in *Drosophila*. In *trp* mutants, maintained light leads to response decay and temporary total loss of sensitivity (inactivation). Using genetically targeted PIP<sub>2</sub>-sensitive inward rectifier channels (Kir2.1) as biosensors, we provide evidence that *trp* decay reflects depletion of PIP<sub>2</sub>. Two independent mutations in the PIP<sub>2</sub> recycling pathway (*rdgB* and *cds*) prevented recovery from inactivation. Abolishing Ca<sup>2+</sup> influx in wild-type photoreceptors mimicked inactivation, while raising Ca<sup>2+</sup> by blocking Na<sup>+</sup>/Ca<sup>2+</sup> exchange prevented inactivation in *trp*. The results suggest that Ca<sup>2+</sup> influx prevents PIP<sub>2</sub> depletion by inhibiting PLC activity and facilitating PIP<sub>2</sub> recycling. Without this feedback one photon appears sufficient to deplete the phosphoinositide pool of ~4 microvilli.

## Introduction

The light sensitive current in *Drosophila* is mediated by two classes of Ca<sup>2+</sup> permeable channels, TRP and TRPL. Both channels are activated downstream of phospholipase C and are the prototypical members of an emerging family of ion channels responsible for a variety of receptor mediated Ca<sup>2+</sup> influx phenomena (Montell and Rubin, 1989; Hardie and Minke, 1992; Phillips et al., 1992; Niemeyer et al., 1996; reviewed in Harteneck et al., 2000). TRP, the major charge carrier, is highly Ca<sup>2+</sup> permeable ( $P_{Ca}:P_{Na} > 100$ ); in *trp* (*transient receptor potential*) mutants lacking this channel, the remaining light induced current (LIC) is carried by TRPL channels (Niemeyer et al., 1996), which have a reduced Ca<sup>2+</sup> permeability ( $P_{Ca}:P_{Na} \approx 4$ ) (Reuss et al., 1997). Ca<sup>2+</sup> influx via these channels is responsible for virtually all the massive (high micromolar) light-induced Ca<sup>2+</sup> rise in *Drosophila* photoreceptors (Peretz et al., 1994; Ranganathan et al., 1994;

Hardie, 1996; Cook and Minke, 1999; Minke and Hardie, 2000).

The *trp* mutation is so named because the response to light decays to baseline during prolonged illumination, following which the photoreceptor becomes temporarily unresponsive (Cosens and Manning, 1969; Minke et al., 1975). Despite over 30 years of investigation, the underlying cause of this phenotype is still controversial. Minke (1982) concluded that the phenotype represented exhaustion of some factor required for excitation. It was later proposed that this factor was Ca<sup>2+</sup>, that Ca<sup>2+</sup> influx via TRP channels was required for refilling Ca<sup>2+</sup> stores and that the decay represented store depletion (Minke and Selinger, 1991). This influential hypothesis led to the proposal that TRP represented a store-operated Ca<sup>2+</sup> influx pathway. While some vertebrate TRP homologues are strong candidates for store-operated channels (review Harteneck et al., 2000), the role of Ca<sup>2+</sup> stores in *Drosophila* phototransduction has become controversial (Acharya et al., 1997; Raghu et al., 2000; Sullivan et al., 2000). Recently, Scott et al. (1997) proposed an alternative explanation for the *trp* phenotype—namely Ca<sup>2+</sup> dependent inactivation of the TRPL channels. In support of this, they reported that the decay was abolished in the absence of external Ca<sup>2+</sup>. These results were challenged by Cook and Minke (1999), who found that responses in *trp* did decay in Ca<sup>2+</sup>-free solutions and that the subsequent response inactivation was more profound.

We have also demonstrated that TRPL channels are subject to Ca<sup>2+</sup>-dependent inactivation (Reuss et al., 1997), but doubted whether this was related to the inactivation following the *trp* decay, since Ca<sup>2+</sup>-dependent inactivation is a normal feature of wild-type photoreceptors (Hardie and Minke, 1994; Hardie and Minke, 1995). However, in view of the questionable role of Ca<sup>2+</sup> stores, we wondered whether depletion of another factor might underlie the *trp* decay. One candidate is phosphatidylinositol 4,5 bisphosphate (PIP<sub>2</sub>), the substrate for PLC, which is the key effector enzyme in *Drosophila* phototransduction (Bloomquist et al., 1988). To test this, we used a genetically targeted PIP<sub>2</sub>-sensitive ion channel as a biosensor to provide evidence that PIP<sub>2</sub> is depleted during the *trp* decay, and showed that mutations in the PIP<sub>2</sub> recycling pathway prevent recovery of sensitivity following inactivation. Our results suggest that Ca<sup>2+</sup> influx normally protects the photoreceptor from PIP<sub>2</sub> depletion by inhibiting PLC and facilitating PIP<sub>2</sub> recycling. The data allow quantitative estimates of the amplification of the transduction cascade and have interesting implications for the strategy of phototransduction.

## Results

### Ca<sup>2+</sup> Dependence of *trp* Phenotype

To gain insight into the *trp* phenotype, we quantified its intensity dependence using whole-cell voltage clamped light responses. In normal Ca<sup>2+</sup>, we found that the light

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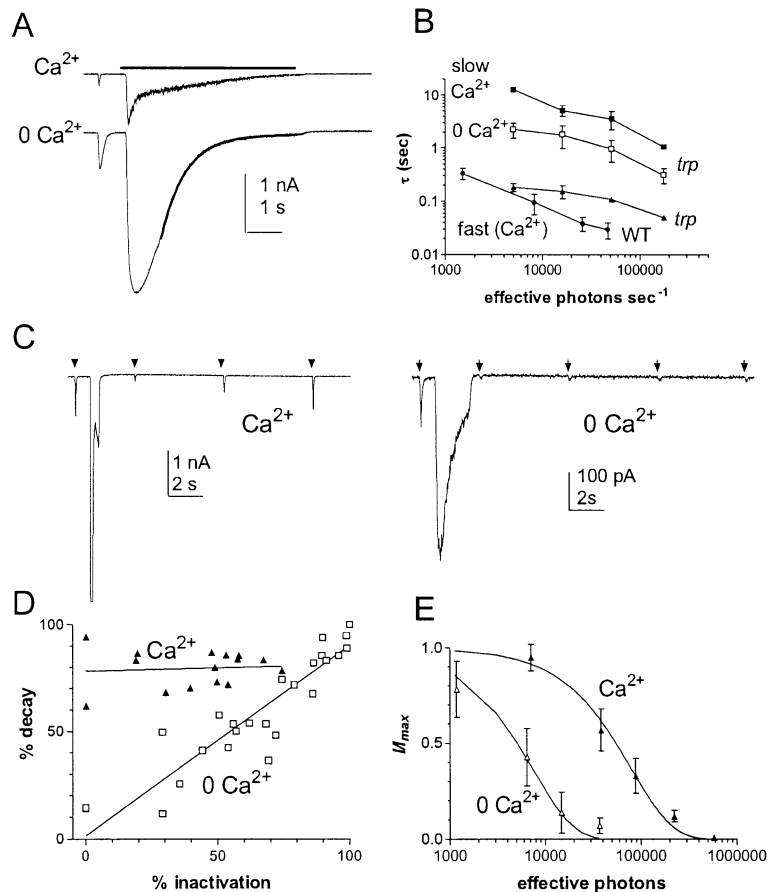


Figure 1. *trp* Decay and Response Inactivation

(A) Responses of *trp* photoreceptors to 5 s light pulses (50,000 effective photons/s) in normal (1.5 mM  $\text{Ca}^{2+}$ ) and  $\text{Ca}^{2+}$ -free bath. In 1.5 mM  $\text{Ca}^{2+}$  (top trace), responses decayed with two kinetic components (see also Figure 3); in  $\text{Ca}^{2+}$  free, responses were facilitated and decayed with a single component. Traces fitted with single ( $\text{Ca}^{2+}$  free) or double exponentials (1.5 mM  $\text{Ca}^{2+}$ ).

(B) Summary of decay time constants (mean  $\pm$  SD,  $n = 5-9$  cells). Closed square, slow component in 1.5 mM  $\text{Ca}^{2+}$ ; closed triangle, fast component, 1.5 mM  $\text{Ca}^{2+}$ ; open square,  $\text{Ca}^{2+}$ -free slow component; closed circle, fast component in wild type (WT).

(C) In zero  $\text{Ca}^{2+}$ , the decay induced by a 2 s stimulus (35,000 effective photons) led to long term response inactivation to dim test flashes (arrows), but in 1.5 mM  $\text{Ca}^{2+}$  (left), despite extensive rapid  $\text{Ca}^{2+}$  dependent decay, a brighter, 500 ms stimulus ( $\sim 10^5$  effective photons, peak response clipped) resulted in little long-term inactivation.

(D) Inactivation (estimated from response to test flash 10 s after the inactivating stimulus) was strongly correlated with decay in  $\text{Ca}^{2+}$ -free solution (open square, linear regression,  $r^2 = 0.84$ ), but not in normal  $\text{Ca}^{2+}$  (closed triangle,  $r^2 = 0.17$ ).

(E) Residual sensitivity ( $I/I_{max}$ ) as a function of effective photon content of inactivating stimulus (closed triangle, 3 s stimuli in 1.5 mM  $\text{Ca}^{2+}$ ; open square, 0.5 s stimuli in  $\text{Ca}^{2+}$ -free bath;  $n = 5-7$  cells). Data fitted assuming each photon inactivates 0.38 microvilli in  $\text{Ca}^{2+}$  and 4.1 microvilli in  $\text{Ca}^{2+}$ -free bath (Equation 2).

induced current (LIC) in *trp* decayed toward baseline with two distinct kinetic components: a rapid (subsecond) decay which appeared similar to, though slightly slower than, the  $\text{Ca}^{2+}$ -dependent inactivation seen in wild-type photoreceptors, and a much slower decay which returned the current to baseline over a period of 1–5 s (Figure 1; see also Figure 3). In most cases the entire decay time course could be well fitted by the sum of two exponentials, although at higher intensities a damped oscillation was often observed in the response waveform (e.g., Figure 1C).

We suspected that the faster component represented the  $\text{Ca}^{2+}$  dependent inactivation of TRPL channels previously described (Reuss et al., 1997; Scott et al., 1997). To test this, we perfused *trp* cells with  $\text{Ca}^{2+}$ -free Ringer's. As previously reported, this immediately resulted in pronounced facilitation due to alleviation of  $\text{Ca}^{2+}$  dependent inactivation (Reuss et al., 1997). In confirmation of Cook and Minke (1999), we found that stimuli that induced decay under normal conditions still induced decay in  $\text{Ca}^{2+}$ -free conditions. However, the fast component was absent and responses now decayed with a single exponential time course,  $\sim 3\times$  faster than the slower component recorded in the presence of  $\text{Ca}^{2+}$  (Figures 1A and 1B).

To test if either component of decay was correlated with response inactivation, we delivered dim test flashes before and after stimuli of various intensities and duration. Only in the absence of  $\text{Ca}^{2+}$  was the response

inactivation (% reduction in response) correlated with the extent of the decay (Figures 1C and 1D). By contrast, in normal  $\text{Ca}^{2+}$ , brief stimuli ( $<1$  s), which induced as much as 80% fast decay, resulted in relatively little loss of sensitivity, which recovered quickly ( $<20$  s), and only longer stimuli that induced slow decay resulted in significant long term inactivation.

We quantified the intensity dependence of response inactivation from similar data by plotting the residual sensitivity ( $S = I/I_{max}$ ) as a function of the intensity of the inactivating stimulus that can be accurately calibrated in terms of effectively absorbed photons (i.e., effectively activated rhodopsin molecules; see Experimental Procedures). Since the number of microvilli ( $m$ ) in the rhabdomere can also be estimated ( $\sim 30,000$ , using morphological data from Suzuki et al., 1993), the fraction ( $f$ ) of microvilli that fail to absorb a photon in a flash of effective photon content  $p$  can be simply calculated from Poisson statistics as

$$f = e^{(-p/m)} \quad (1)$$

Assuming that the residual sensitivity is determined by the fraction of microvilli remaining unaffected by a given stimulus, it is then possible to express inactivation in terms of the number of microvilli inactivated by each effectively absorbed photon. Namely, if one photon inactivates  $n$  microvilli, the residual sensitivity is predicted by

Table 1. Response Inactivation

	1.5 mM Ca <sup>2+</sup>	0 Ca <sup>2+</sup>
Wild type	NA	1.7 ± 1.1 (9)
<i>trp</i> <sup>343</sup>	0.42 ± 0.13 (6)	4.2 ± 1.5 (6)
<i>rdgB</i> <sup>K<sup>S</sup>222</sup>	0.39 ± 0.16 (4)	3.1 ± 0.7 (9)
<i>cds</i> <sup>1</sup>	1.0 ± 0.3 (2)	4.7 ± 1.4 (3)

Number of microvilli inactivated by each effectively absorbed photon under different conditions leading to long-term response inactivation attributable to PIP<sub>2</sub> depletion. Data estimated from the residual sensitivity 10 s after inactivating stimuli of various intensities from Equations 2 and 3, expressed as mean ± SD (number of cells).

$$S = e^{(-p/(m/n))} \quad (2)$$

from which

$$n = -\ln S \times (m/p) \quad (3)$$

In the presence of Ca<sup>2+</sup>, brief stimuli (0.5 s) failed to induce any significant inactivation in *trp* beyond wild-type controls (not shown); with stimuli of long duration (3 s), the loss of sensitivity was fitted assuming each photon effectively inactivated ~0.4 microvilli. In zero Ca<sup>2+</sup>, inactivation was independent of stimulus duration (between 0.5 and 5s), but enhanced 10-fold, each photon effectively inactivating ~4 microvilli (Figure 1E; Table 1).

#### Kir2.1 Channels as Biosensors of PIP<sub>2</sub>

Because we doubted whether the *trp* phenotype reflected Ca<sup>2+</sup> store depletion, we sought evidence for an alternative explanation, namely, depletion of PIP<sub>2</sub>—the substrate for the key effector enzyme in phototransduction. Support for this might be provided by biochemical measurements of microvillar PIP<sub>2</sub>, but such measurements would be technically challenging and difficult to relate to the *in vivo trp* decay. We therefore developed a strategy for monitoring PIP<sub>2</sub> dynamics *in vivo*. Specifically, we exploited the properties of the Kir2.1 inward-rectifier channel, the activity of which is believed to be primarily determined by binding to PIP<sub>2</sub> (Huang et al., 1998; Rohacs et al., 1999). We expressed the human Kir2.1 channel in *Drosophila* photoreceptors under control of the rhodopsin (Rh1) promoter (see Experimental Procedures). The Kir2.1 transgene included an inframe EGFP tag, and predominant or exclusive expression in the microvillar membrane was confirmed by observation of fluorescence in both the intact eye (Figure 2) and dissociated ommatidia. All cells (n > 100) expressing Kir2.1 channels displayed a large constitutive potassium conductance immediately on establishing the whole-cell configuration, but light responses were unaffected. The conductance (IRK) showed the well-known inwardly rectifying properties of Kir2.1 as well as its characteristic voltage dependent block by external Cs<sup>+</sup> (Figure 2). Under control conditions, the IRK current was maintained for the lifetime of the recording (up to 20 min) as long as ATP was included in the pipette. In the absence of ATP, the current gradually decayed, as is typical for the behavior of Kir channels and normally attributed to degradation of PIP<sub>2</sub> by lipid phosphatases (Huang et al., 1998).

Following stimulation by light under control conditions, the IRK current showed at most a slight suppres-

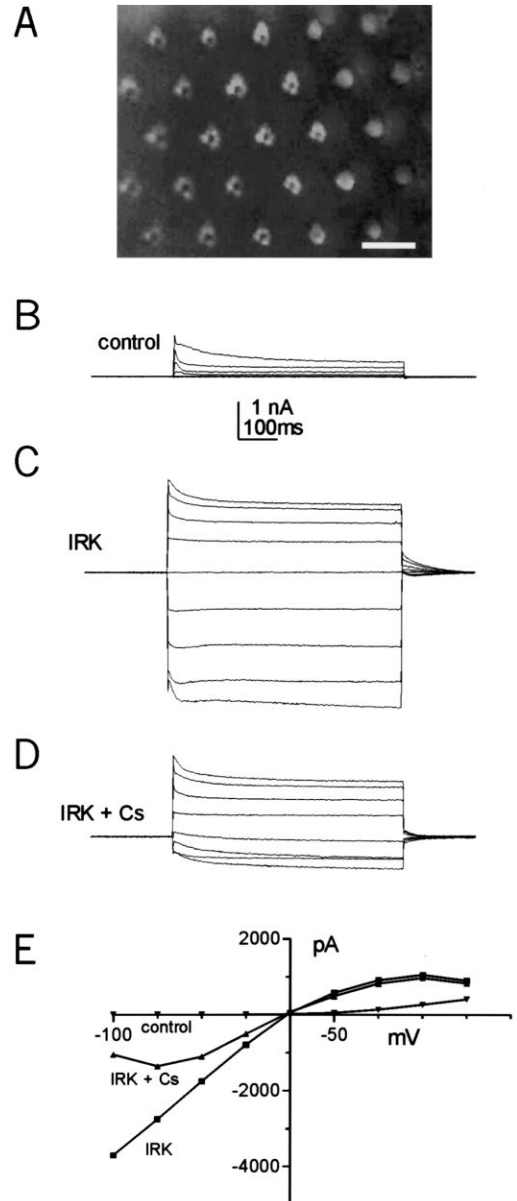


Figure 2. Functional Expression of Kir2.1 in *Drosophila* Photoreceptors

(A) Fluorescence of *Drosophila* rhabdomeres in Kir2.1-EGFP expressing flies observed in the intact eye using optical neutralization of the cornea (see Methods). The central rhabdomeres (R7 and R8) appeared dark, since Rh1GAL4 drives expression only in R1–6 cells. Scale bar 20 μm.

(B–D) Voltage clamped currents evoked by voltage steps from 0 mV to –100 mV (10 mV steps) from a holding potential of –70 mV in a control (B: Rh1GAL4/+ heterozygote), and Kir2.1 expressing photoreceptor (C and D). Bath contained 10 mM K<sup>+</sup>. In control cells, outward currents representing voltage gated outward currents (I<sub>A</sub> and delayed rectifier) were elicited at voltages above ~–50 mV; in Kir2.1-expressing cells these are superimposed on a large IRK, which was partially blocked in a voltage dependent manner by 4 mM Cs (D).

(E) I–V traces of currents in (B–D) determined at end of the voltage steps (open triangle, control; closed square, IRK). Note the voltage-dependent block in the presence of Cs<sup>+</sup> (closed triangle).

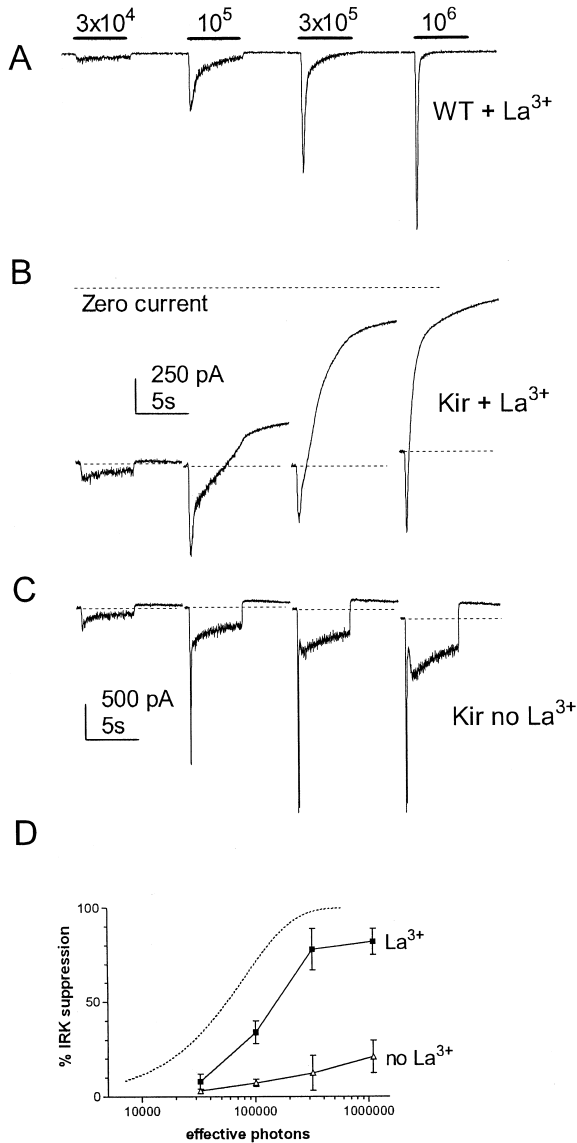


Figure 3. *trp* Decay Correlates with IRK Suppression

(A and B) Responses to 5 s stimuli (bars) of increasing intensity (number of effectively absorbed photons indicated) in bath containing 40  $\mu$ M La<sup>3+</sup> to mimic the *trp* phenotype.

(A) In a control cell, the lowest intensity evoked a maintained response, but a partial decay of the LIC was apparent with the second flash (10<sup>5</sup> photons) and decay was essentially complete with the two highest intensities.

(B) Similarly, significant IRK suppression was first seen with 10<sup>5</sup> photons and was nearly complete at the two higher intensities. The constitutive IRK current in the dark (dotted lines) was  $-1.3$  nA; zero current level also shown by a dotted line.

(C) Responses in a Kir2.1 expressing cell in normal bath (no La<sup>3+</sup>) show the typical peak/plateau relation of the LIC but only slight suppression of IRK (constitutive current  $\sim -1$  nA).

(D) Intensity dependence of IRK suppression for 5 s flashes in the presence (closed square) and absence (open triangle) of La<sup>3+</sup>. Mean  $\pm$  SD,  $n = 4-7$  cells for each data point. Dotted line: intensity dependence of *trp* response inactivation ( $100 \times [1 - I/I_{max}]$ , from Figure 1E).

sion (maximally  $\sim 20\%$ , Figure 3C). This may represent loss of PIP<sub>2</sub>, but we cannot exclude some modulation of the current by the large ion fluxes associated with

the LIC. In order to mimic the *trp* phenotype, the photoreceptors were exposed to La<sup>3+</sup> (40  $\mu$ M), which completely blocks the TRP channels while leaving TRPL channels unaffected, and quantitatively mimics diverse aspects of the physiological *trp* phenotype (e.g., Hochstrate, 1989; Hardie and Minke, 1992). Under these conditions, the IRK current was now profoundly suppressed with an intensity dependence that closely matched that of *trp* decay and response inactivation (Figures 3B and 3D). Light-induced ion fluxes are substantially reduced under these conditions, due to the block of TRP channels, leaving PIP<sub>2</sub> depletion as the most obvious explanation for the suppression of IRK. Following suppression, the IRK current recovered to preillumination levels over a period of  $\sim 1$  min ( $t_{1/2} = 32 \pm 5$  s,  $n = 11$ ; see, e.g., Figure 4), presumably reflecting PIP<sub>2</sub> resynthesis.

#### Mutations in the PIP<sub>2</sub> Recycling Pathway Prevent Recovery from Inactivation

PIP<sub>2</sub> is resynthesized by conversion of DAG to phosphatidic acid, synthesis of phosphatidylinositol (PI) via CDP-diacylglycerol, transport of PI to plasma membrane, and serial phosphorylation to PIP<sub>2</sub> (Batty et al., 1998). If *trp* decay and inactivation are due to PIP<sub>2</sub> depletion, then recovery from inactivation should be impaired in mutants of this pathway. We examined two such mutants, namely *rdgB*<sup>K5222</sup>, a severe hypomorph of PI transfer protein (PITP) (Vihtelic et al., 1993), and *cds*<sup>1</sup>, which lacks CDP-diacylglycerol synthase (Wu et al., 1995). As long as flies were strictly dark reared, we found that responses to moderate flashes in *rdgB* were indistinguishable from wild type under control conditions. In *cds*, peak amplitudes were slightly reduced ( $\sim 50\%$  of wild-type controls) and time-to-peaks slightly delayed ( $\sim 70$  ms cf. 50 ms in wild type), probably indicative of reduced PIP<sub>2</sub> levels. Quantum bumps were indistinguishable from wild type in both mutants (data not shown).

To mimic the *trp* phenotype, *rdgB* or *cds* photoreceptors were exposed to La<sup>3+</sup>. A dim flash was delivered to test sensitivity, following which cells were challenged with a single 5 s stimulus of intensity just sufficient to induce full decay. While *trp* photoreceptors (or wild type in the presence of La<sup>3+</sup>) fully recovered sensitivity from such flashes within  $\sim 90$  s, in *rdgB* ( $n = 7$ ) and *cds* ( $n = 5$ ), no recovery was seen for the duration of the experiment (Figure 4). Interestingly, in most cases (12/15), similar stimuli delivered to *rdgB* in the absence of La<sup>3+</sup> resulted in responses which failed to terminate, leaving channels constitutively activated (Figure 4C). Similar behavior has also been reported in electroretinogram recordings (Milligan et al., 1997). The lack of any recovery in *rdgB* or *cds* may seem surprising because PI and PIP kinases are presumably still present and capable of synthesizing PIP<sub>2</sub> from PI in the microvilli. However, these kinases probably operate too quickly for their effect to be noticed, i.e., PI is converted to PIP<sub>2</sub> "on demand" during the response. This implies not only all the PIP<sub>2</sub>, but also all microvillar PI was depleted by the inactivating flashes.

From in vitro studies and by sequence homology, the *rdgB* protein is assumed to function as a PITP and is thus expected to be an essential component of the PI recycling pathway (Vihtelic et al., 1993; Milligan et al.,

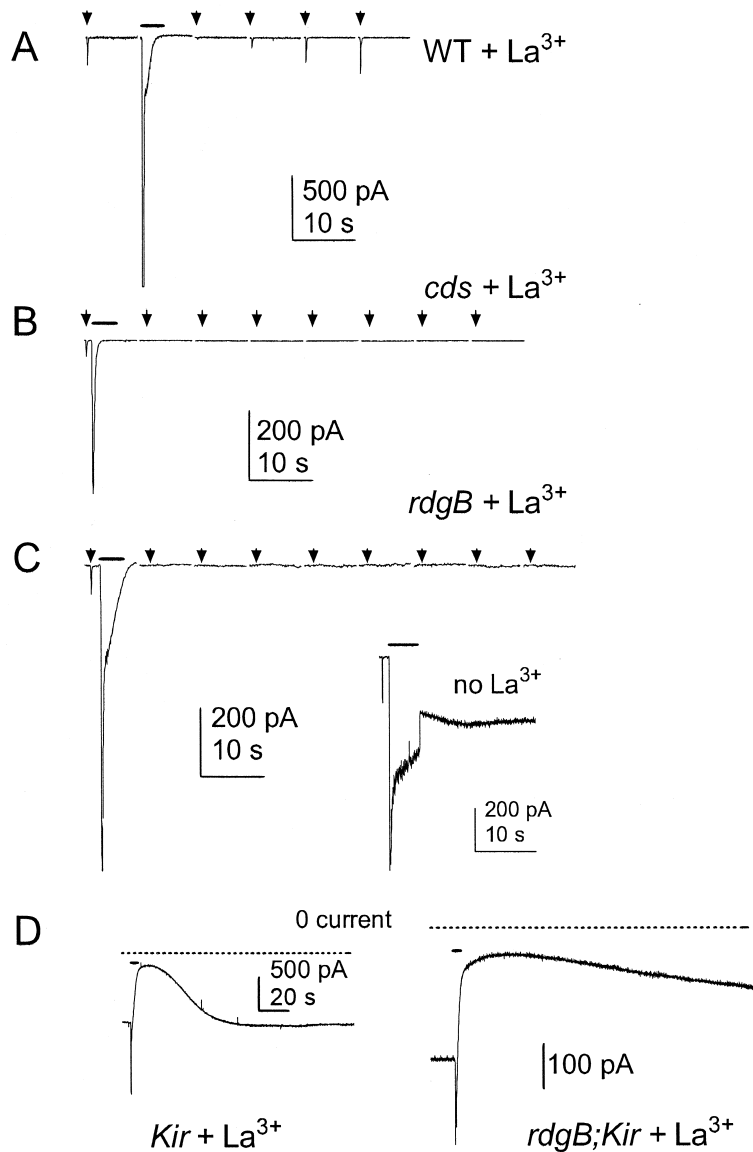


Figure 4. Mutations in PIP<sub>2</sub> Recycling Pathway Prevent Recovery from Response Inactivation

(A) In a wild-type (WT) cell bathed in 40 μM La<sup>3+</sup>, a stimulus of intensity and duration just sufficient to induce complete decay (5 s, ~200,000 photons) inactivated the response to a dim test flash (500 photons, arrows), which subsequently recovered over ~1 min (20 s between start of each trace).

(B and C) In photoreceptors from *rdgB* or *cds* mutants exposed to La<sup>3+</sup>, similar stimuli also led to decay; but there was no recovery for the duration of the experiment (30 s between traces).

(C) Inset: response to same stimulus in *rdgB* without La<sup>3+</sup> failed to terminate.

(D) Suppression of IRK current in photoreceptors expressing Kir2.1 channels, exposed to La<sup>3+</sup> and stimulated with a 5 s inactivating flash. Left: in an otherwise wild-type fly, IRK was suppressed by ~70%, recovering within 60 s. Right: on an *rdgB* background, recovery from suppression was nearly abolished.

1997). However, there is no *in vivo* evidence for this assumption. To test whether PI recycling was in fact impaired, we generated *rdgB* mutant lines expressing Kir2.1 channels. After the light-induced suppression in the presence of La<sup>3+</sup>, the IRK current in otherwise wild-type flies fully recovered with a half-time of ~30 s (see above). In dark-reared *rdgB;Kir* flies after one flash of intensity and duration just sufficient to induce decay, the IRK current was similarly suppressed but then recovered by at most ~50% over a period of several minutes (n = 6, Figure 4D), probably reflecting some limited, slow transfer of PI by residual RDGB protein in this hypomorph.

#### Response Inactivation in Ca<sup>2+</sup>-Free Solutions

Because the primary defect in *trp* mutants is the absence of a Ca<sup>2+</sup> permeable channel, the simplest explanation for the various manifestations of the *trp* phenotype would be the reduced Ca<sup>2+</sup> influx, in which case

removing external Ca<sup>2+</sup> might be expected to mimic the *trp* phenotype. Although prolonged stimuli delivered to wild-type cells in Ca<sup>2+</sup>-free solution do not normally lead to response decay unless cells have been exposed to Ca<sup>2+</sup> for long periods (Hardie and Minke, 1992), this might reflect differences between TRP and TRPL channels, such as their relative sensitivity to putative second-messenger molecules or mechanism of inactivation. Therefore, we asked, instead, whether the *trp* response inactivation phenotype was mimicked in Ca<sup>2+</sup>-free solutions. Sensitivity in wild type normally recovers to ~80% of control levels within seconds of even the brightest "inactivating" stimuli; however, in the absence of Ca<sup>2+</sup>, sensitivity was progressively reduced with increasing intensity, sensitivity recovering slowly with a time course (t<sub>1/2</sub> = 47 ± 10 s, n = 6; Figure 5B) very similar to that following response inactivation in *trp* or wild type in the presence of La<sup>3+</sup> (t<sub>1/2</sub> = 52 ± 6 s, n = 7; e.g., Figure 4A). With inactivating flashes containing more than ~20,000

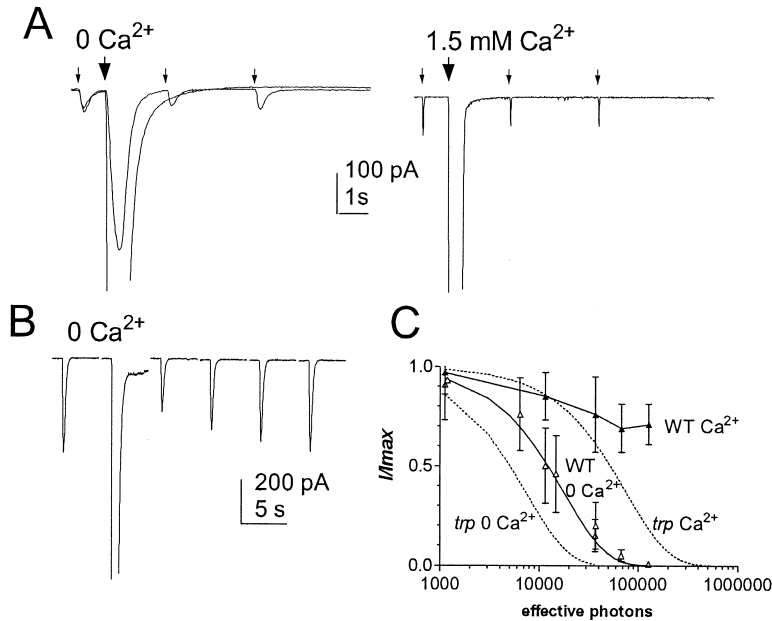


Figure 5. Response Inactivation in  $\text{Ca}^{2+}$ -Free Ringer's

Sensitivity in a wild-type photoreceptor was probed with dim test flashes (small arrows) delivered before and after an inactivating stimulus of either 1000 or 35,000 photons.

(A) The photoreceptor was completely inactivated following the brighter stimulus in  $0 \text{ Ca}^{2+}$  (left) but not in  $1.5 \text{ mM Ca}^{2+}$  (right).

(B) Time course of recovery from inactivation in  $\text{Ca}^{2+}$ -free solution following an inactivating flash containing  $\sim 26,000$  photons: 28 s between traces.

(C) The residual sensitivity ( $I/I_{\text{max}}$ ) in  $0 \text{ Ca}^{2+}$  (open triangle) was fitted assuming each effectively absorbed photon inactivated 1.7 microvilli (Equation 2); closed triangle data obtained in an identical fashion in normal  $\text{Ca}^{2+}$  (mean  $\pm$  SD,  $n = 4-5$  cells). Dotted lines: equivalent data from *trp* photoreceptors (from Figure 1E).

effective photons, responses often failed to terminate, leaving channels constitutively activated, sometimes indefinitely, reminiscent of the behavior of *rdgB* photoreceptors in the presence of  $\text{Ca}^{2+}$ . Quantitatively, the number of microvilli (1.7) effectively inactivated per photon in  $\text{Ca}^{2+}$ -free solutions was  $\sim 4\times$  greater than in *trp* in normal  $\text{Ca}^{2+}$  and within a factor of  $\sim 2$  of that measured in *trp* in  $\text{Ca}^{2+}$ -free solutions (Figure 5; Table 1).

To test whether  $\text{PIP}_2$  depletion was also responsible for inactivation in zero  $\text{Ca}^{2+}$ , we again recorded from *rdgB* and *cds* photoreceptors. Because responses in *rdgB*, in particular, often failed to terminate with brighter flashes, we used relatively dim flashes repeated at 30 s intervals in  $\text{Ca}^{2+}$ -free solutions. In wild-type controls, this was sufficient time for the response to recover fully between flashes; however, in *rdgB* and *cds*, each flash induced an exponential and irreversible reduction in sensitivity (Figure 6; see also Wu et al., 1995). The sensi-

tivity loss was well fitted, assuming each photon inactivated  $\sim 3$  microvilli. Similar experiments performed in the presence of  $\text{Ca}^{2+}$  also led to a progressive irreversible loss of sensitivity, but  $\sim 8\times$  as many photons were required to achieve the same degree of inactivation (Table 1), implying that  $8\times$  less  $\text{PIP}_2$  was hydrolyzed per absorbed photon. These results show that removing  $\text{Ca}^{2+}$  closely mimics the *trp* inactivation phenotype, and that this is also likely to be due to depletion of  $\text{PIP}_2$ . The close quantitative correspondence between response inactivation measured in *trp* and *rdgB* (Table 1) raises the possibility that the *trp* phenotype may reflect a defect in  $\text{PIP}_2$  recycling (see Discussion).

#### $\text{Ca}^{2+}$ Rescues the *trp* Inactivation Phenotype

It has been suggested that reduced  $\text{Ca}^{2+}$  influx is not sufficient to explain the *trp* phenotype because it could not be rescued by raising extracellular or intracellular

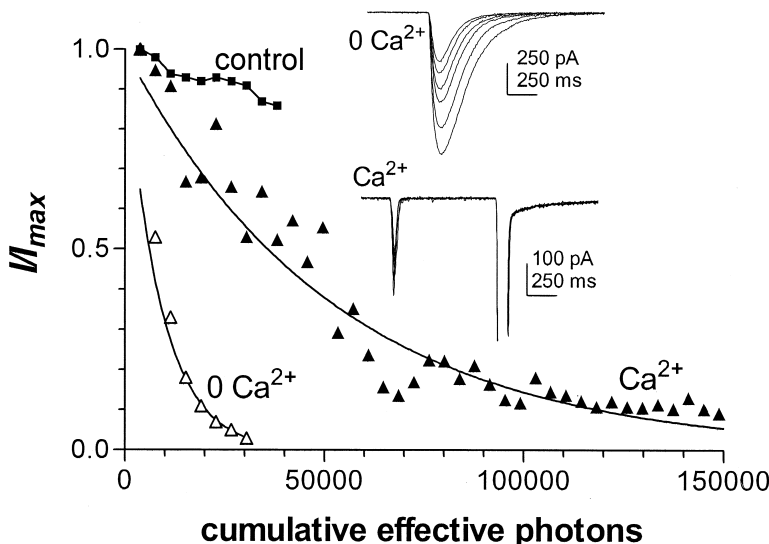


Figure 6. Response Inactivation in *rdgB*

Insets: responses to 10 ms flashes repeated at 30 s intervals in an *rdgB* photoreceptor exposed to  $\text{Ca}^{2+}$ -free solution: each flash ( $\sim 2000$  photons) induced a progressive reduction in sensitivity. In  $1.5 \text{ mM Ca}^{2+}$  repeated flashes of twice the intensity induced significantly less inactivation (sensitivity probed with a dim test flash before each flash as the large responses to the inactivating stimulus were inadequately clamped). The plot of normalized peak responses ( $I/I_{\text{max}}$ ) confirms that loss of sensitivity was more severe in  $\text{Ca}^{2+}$ -free bath (open triangle) than in  $1.5 \text{ mM Ca}^{2+}$  (closed triangle); wild-type controls in  $\text{Ca}^{2+}$ -free bath (closed square) showed little inactivation under these conditions. Curves were fitted assuming that each absorbed photon inactivates 3.4 microvilli in  $\text{Ca}^{2+}$  free but only 0.58 microvilli in the presence of  $\text{Ca}^{2+}$  (Equation 2). See Table 1 for statistics on all cells.

Ca<sup>2+</sup> (Scott et al., 1997; Cook and Minke, 1999). However, it may not be possible to reproduce wild-type microvillar Ca<sup>2+</sup> levels in *trp* mutants by raising extracellular Ca<sup>2+</sup> because their light-sensitive channels (i.e., TRPL channels) are ~25× less permeable to Ca<sup>2+</sup> (Reuss et al., 1997), and quantum bumps in *trp* are ~5× smaller (Niemeyer et al., 1996; Henderson et al., 2000). A potentially more powerful method to raise Ca<sup>2+</sup> is to block Na<sup>+</sup>/Ca<sup>2+</sup> exchange during the response. This is because, following stimulation, Ca<sup>2+</sup> initially rises to very high (>200 μM) concentrations in the microvilli, but then relaxes quickly (~100 ms) to a much lower steady-state value, due to Na<sup>+</sup>/Ca<sup>2+</sup> exchange (Oberwinkler and Stavenga, 2000). A similar behavior can be expected in *trp*, except the absolute levels reached will be lower due to the reduced Ca<sup>2+</sup> influx. We reasoned that in *trp*, the transient Ca<sup>2+</sup> rise might be sufficient to prevent PIP<sub>2</sub> depletion, before falling below some critical value as it is extruded by the exchanger. If this is the case, blocking the exchanger should maintain Ca<sup>2+</sup> at sufficient levels to prevent depletion. We therefore rapidly perfused photoreceptors with a solution in which Na<sup>+</sup> was substituted for Cs<sup>+</sup> or Li<sup>+</sup> to prevent Na<sup>+</sup>/Ca<sup>2+</sup> exchange, starting shortly (<1 s) before a flash, which would normally inactivate the response in *trp*. The degree of inactivation was probed with a test flash 10 s after the inactivating stimulus, by which time reperfusion of the bath solution had restored exchanger activity and presumably returned microvillar Ca<sup>2+</sup> near to preillumination levels.

In controls, the response to the test stimulus in *trp* was virtually abolished by the inactivating stimulus as usual, but if Na<sup>+</sup>/Ca<sup>2+</sup> exchange was blocked during the 5 s, inactivating stimulus sensitivity recovered to ~50% (Figure 7). Responses recorded in the absence of Na<sup>+</sup> decayed to baseline more quickly than under control conditions. We presume this represents enhanced Ca<sup>2+</sup>-dependent inactivation, but it was clearly no longer related to the response inactivation and cannot be considered part of the *trp* phenotype because a similar decay was seen in wild-type flies under these conditions (Figure 7C).

To test whether PIP<sub>2</sub> depletion was prevented under these conditions, we performed similar experiments in flies expressing Kir2.1 channels in the presence of La<sup>3+</sup>. As shown above, a stimulus of sufficient intensity and duration to induce decay resulted in ~70% suppression of the IRK current indicating substantial loss of PIP<sub>2</sub>. However, when Na<sup>+</sup>/Ca<sup>2+</sup> exchange was blocked during the inactivating stimulus, IRK was suppressed by at most 10%–20% (Figure 7D).

## Discussion

We have shown that responses in *trp* decay with two kinetic components, but only the slower component, which was accelerated in Ca<sup>2+</sup>-free solution, was correlated with response inactivation. By targeting a PIP<sub>2</sub>-sensitive ion channel to the microvilli, we provided evidence that decay was associated with depletion of PIP<sub>2</sub>, suggesting this may underlie the *trp* phenotype. This was supported by two independent mutations in the PIP<sub>2</sub> recycling pathway, both of which prevented recovery from inactivation. Finally we demonstrated that re-

moving extracellular Ca<sup>2+</sup> mimicked response inactivation, while raising Ca<sup>2+</sup> could rescue this aspect of the *trp* phenotype. In the following, we discuss the use of Kir channels as PIP<sub>2</sub> biosensors, compare our findings with previous data and models of *trp* decay, and discuss novel aspects of the Ca<sup>2+</sup>-dependent regulation of PIP<sub>2</sub> metabolism revealed by this study. Finally, we consider the implications of our results for the cellular and molecular strategies underlying the performance of microvillar photoreceptors.

## Kir Channels as PIP<sub>2</sub> Biosensors

Only recently have attempts been made to monitor PIP<sub>2</sub> in vivo, in particular using GFP-tagged pleckstrin homology (PH) domains (Stauffer et al., 1998; Varnai and Balla, 1998; Holz et al., 2000). Although this approach has proved valuable for visualizing PIP<sub>2</sub> localization, the PH domain used in most of these studies (from PLCδ<sub>1</sub>) has higher affinity for InsP<sub>3</sub> than PIP<sub>2</sub>, complicating interpretation of dynamic changes (Lemmon et al., 1995; Hirose et al., 1999). We therefore developed an alternative approach for tracking PIP<sub>2</sub>, by exploiting the properties of the Kir family of inward rectifier K channels, whose activity has been shown to depend upon binding to PIP<sub>2</sub> (Baukowitz et al., 1998; Huang et al., 1998; Zhang et al., 1999). We chose Kir2.1 (= IRK1; Kubo et al., 1993), as this is constitutively active, is activated by PIP<sub>2</sub> alone without cofactors, and is reported to have the highest affinity and greatest specificity for PIP<sub>2</sub> of all the Kir family (Huang et al., 1998; Rohacs et al., 1999; Zhang et al., 1999).

Although there are no quantitative dose response data for the PIP<sub>2</sub> dependence of Kir channels, it seems likely that the near total suppression seen in the presence of La<sup>3+</sup> also represents near total depletion of microvillar PIP<sub>2</sub> and PI. Thus, firstly, the suppression of IRK currents was invariably associated with profound loss of sensitivity to light, entirely consistent with loss of substrate (PIP<sub>2</sub>) for phototransduction. Secondly, Kir2.1's affinity for PIP<sub>2</sub> is so high that it has not previously proven possible to suppress its activity by PLC activation (Kobrinisky et al., 2000). The high affinity for PIP<sub>2</sub> may also explain why IRK required slightly higher intensities for suppression than *trp* inactivation (Figure 3D), recovered slightly more quickly (t<sub>1/2</sub> = 32 s cf. ~50 s for recovery of light response), and, unlike the response to light, showed some limited slow recovery in *rdgB* hypomorphs.

## Ca<sup>2+</sup>-Dependent Regulation of PIP<sub>2</sub>

Our results indicate that the stimulus-dependent loss of sensitivity when Ca<sup>2+</sup> influx is reduced—either by removing extracellular Ca<sup>2+</sup> or by the *trp* mutation—is due to PIP<sub>2</sub> depletion. Assuming PIP<sub>2</sub> levels are determined by the balance between synthesis and hydrolysis, this could be explained by Ca<sup>2+</sup>-dependent inhibition of PLC activity, or by Ca<sup>2+</sup>-dependent facilitation of PIP<sub>2</sub> recycling. Our evidence suggests both may contribute. Firstly, facilitation of PIP<sub>2</sub> recycling cannot readily account for the results found in PIP<sub>2</sub>-recycling mutants. Inactivation in *rdgB* and *cds* was irreversible on the time scale of the experiments, indicating there was no effective PIP<sub>2</sub> recycling, and we therefore attribute the

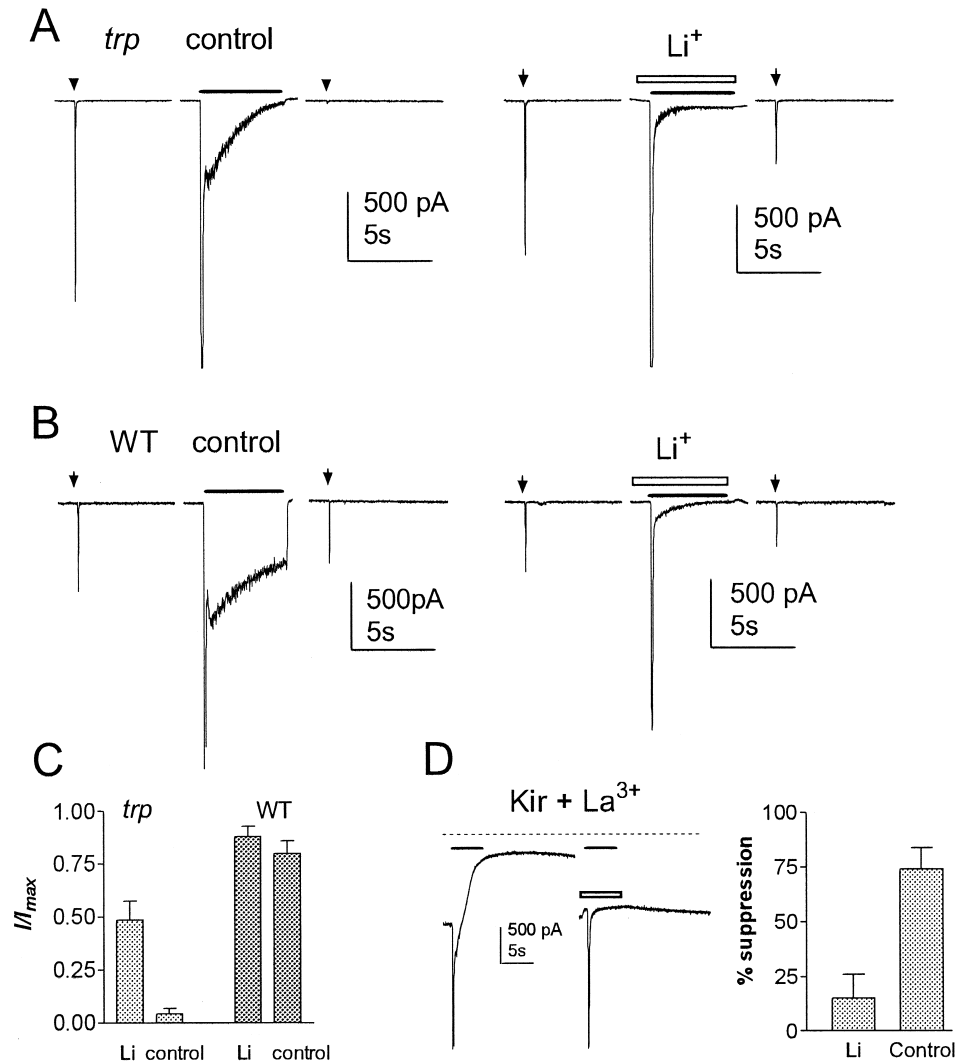


Figure 7. Blocking  $\text{Na}^+/\text{Ca}^{2+}$  Exchange Rescues Response Inactivation in *trp*

In each set of three traces, a test flash preceded a 5 s inactivating stimulus (solid line:  $\sim 50,000$  photons  $\text{s}^{-1}$ ); sensitivity was probed with an identical test flash delivered 10 s later.

(A) Under control conditions (left), the response to a test flash was reduced to  $<5\%$ . However, when the same cell was perfused with  $\text{Na}^+$ -free Ringer (box) during the inactivating stimulus, the response had recovered to  $\sim 50\%$ .

(B) Identical experiment performed in wild type (WT), the response recovered to  $>50\%$  irrespective of perfusion. In both wild-type and *trp* blocking the exchanger resulted in complete decay of the response due to enhanced  $\text{Ca}^{2+}$ -dependent inactivation.

(C) Histogram showing recovery in *trp* (mean  $\pm$  SD,  $n = 6$ ) and WT under control conditions with and without (control)  $\text{Li}^+$  perfusion during the stimulus.

(D) Similar experiment performed in flies expressing Kir2.1 channels exposed to  $\text{La}^{3+}$ . Under control conditions the IRK current was suppressed by the inactivating stimulus; this was prevented by  $\text{Na}^+$ -free perfusion (right). The histogram compares percent suppression of IRK with and without  $\text{Li}^+$  perfusion (mean  $\pm$  SD,  $n = 11$ ).

marked  $\text{Ca}^{2+}$  dependence of inactivation in these mutants to  $\text{Ca}^{2+}$ -dependent inhibition of PLC activity. Because response inactivation in *trp* and *rdgB* showed similar sensitivity to  $\text{Ca}^{2+}$  (Figures 1E and 6; Table 1), the same mechanism—i.e., inhibition of PLC activity—seems likely to have been responsible. This suggests that  $\text{Ca}^{2+}$  influx may also be required to facilitate rapid  $\text{PIP}_2$  recycling and that failure of this may be the primary cause of the *trp* phenotype in the presence of  $\text{Ca}^{2+}$ . This is supported by the close quantitative correspondence between the *rdgB* and *trp* response inactivation phenotypes in both  $\text{Ca}^{2+}$  and  $\text{Ca}^{2+}$ -free solutions (Table 1).

Although the response in *rdgB* does not normally decay to baseline as in *trp*, as in the case of zero  $\text{Ca}^{2+}$ , we assume this reflects the different properties of TRP and TRPL channels. Interestingly, we found that stimuli of sufficient intensity and duration to induce decay in *trp* often resulted in failure of response termination in *rdgB* or in wild type in  $\text{Ca}^{2+}$ -free solutions. This suggests that, although both TRP and TRPL channels require  $\text{PIP}_2$  hydrolysis for activation, under some conditions, TRP channels may become constitutively activated following  $\text{PIP}_2$  depletion, while TRPL channels inactivate.

Both  $\text{Ca}^{2+}$ -dependent inhibition of PLC activity and

Ca<sup>2+</sup>-dependent facilitation of PIP<sub>2</sub> recycling appear required to explain the Ca<sup>2+</sup>-dependent regulation of PIP<sub>2</sub> metabolism indicated by this study. In principle, PLC activity could be regulated by Ca<sup>2+</sup> at the level of rhodopsin, G-protein, or PLC. G-proteins are not normally regulated by Ca<sup>2+</sup>, and recent evidence suggests Ca<sup>2+</sup> is probably not involved in regulating rhodopsin lifetime in *Drosophila* (Alloway and Dolph, 1999). Therefore, PLC itself should be considered as a likely site of action, and indeed *Drosophila* PLC has been reported to be inhibited in vitro by high micromolar concentrations of Ca<sup>2+</sup> (Running Deer et al., 1995). Possible mechanisms for Ca<sup>2+</sup> dependent inhibition of PLC include Ca-calmodulin binding or phosphorylation by a Ca<sup>2+</sup>-dependent enzyme such as PKC, both of which have been proposed in *Limulus* photoreceptors (Richard and Lisman, 1997; Dabdoub and Payne, 1999). However, in view of the exceptionally high Ca<sup>2+</sup> concentrations believed to occur in the microvilli following even a single photon absorption, a simple and direct mechanism might be binding of Ca<sup>2+</sup> to PIP<sub>2</sub>, thereby screening its negative charges.

To our knowledge, there is no precedent for Ca<sup>2+</sup> dependent facilitation of PIP<sub>2</sub> recycling. However, an acidic amino-acid domain adjacent to the PI transfer domain of the RDGB protein has been reported to bind Ca<sup>2+</sup> in vitro (Vihtelic et al., 1993) suggesting a possible molecular target. Because several steps of the PIP<sub>2</sub> recycling pathway are presumed to occur in the submicrovillar cisternae (specialized endoplasmic reticulum at the base of the microvilli), which are also believed to represent Ca<sup>2+</sup> stores, we would not exclude the possibility that PIP<sub>2</sub> recycling might be regulated by store Ca<sup>2+</sup> rather than cytosolic Ca<sup>2+</sup>.

#### Resolution of the *trp* Phenotype

Our results support the proposal that exhaustion of some factor required for excitation underlies the *trp* phenotype (Minke, 1982). We have now identified this factor as PIP<sub>2</sub>, while we found that the role of Ca<sup>2+</sup> is indirect, namely to reduce net PIP<sub>2</sub> hydrolysis. Earlier, apparently contradictory studies of *trp* decay can readily be reconciled with these findings. We confirm that response decay and inactivation in *trp* are more profound in zero Ca<sup>2+</sup> (Cook and Minke, 1999), but explain this by proposing that Ca<sup>2+</sup> influx during the light response in *trp* is still sufficient to inhibit PLC, but insufficient to facilitate rapid PIP<sub>2</sub> recycling. Scott et al.'s report (1997) of Ca<sup>2+</sup>-dependent inactivation of TRPL channels is revealed as the first and faster component of *trp* decay in the presence of Ca<sup>2+</sup>. However, this is unrelated to the response inactivation and cannot be considered part of the mutant phenotype since Ca<sup>2+</sup> dependent inactivation is a characteristic of wild-type photoreceptors as well.

The proposal that *trp* decay represented depletion of Ca<sup>2+</sup> stores (Minke and Selinger, 1991; Cook and Minke, 1999) has been a key argument for the hypothesis that release of Ca<sup>2+</sup> is essential for excitation in *Drosophila* photoreceptors. Together with recent results showing that phototransduction is unaffected by mutations in either the InsP<sub>3</sub> or ryanodine receptors (Acharya et al., 1997; Raghu et al., 2000; Sullivan et al., 2000), our expla-

nation for the *trp* phenotype casts doubt on any direct role for Ca<sup>2+</sup> stores in excitation in *Drosophila* photoreceptors.

#### Quantitative Considerations

To our knowledge, the only available data on the biochemical amplification of the phototransduction cascade in invertebrate photoreceptors come from studies in the squid, suggesting ~500 PIP<sub>2</sub> molecules are hydrolyzed per activated rhodopsin (Szuts, 1993). From the relationship between the intensity of an inactivating flash and sensitivity loss, it is now possible to make an estimate of the biochemical amplification in *Drosophila*. Without Ca<sup>2+</sup> influx or PIP<sub>2</sub> recycling, activation of one rhodopsin molecule appears to result in the hydrolysis, not only of the immediately available PIP<sub>2</sub>, but also most of the PI in up to ~4 microvilli (Table 1). Data from *Calliphora* (Zinkler et al., 1985) suggest that PI represents ~3% of microvillar lipids which, given the dimensions of the microvilli (1 μm long, 60 nm in diameter), would amount to ~7500 molecules per microvillus. The comparison of response inactivation in the presence and absence of Ca<sup>2+</sup> suggests that ~10-fold less (i.e., up to ~3000 PIP<sub>2</sub> molecules) may be hydrolyzed per photon under physiological conditions in the presence of Ca<sup>2+</sup> influx. This would probably represent all the immediately available PIP<sub>2</sub>, which is not generally considered to account for more than 10% of total PI.

#### Strategy of Phototransduction in Microvillar Photoreceptors

Dipteran photoreceptors have the fastest G-protein coupled signaling pathway known: when dark adapted they respond to single photons with large bumps (~10 pA in *Drosophila*), representing simultaneous activation of about 15 TRP channels (Henderson et al., 2000) with a duration of ~30 ms at 20°C. They also light adapt to cover the entire environmental range of intensities (up to ~10<sup>5</sup>-10<sup>6</sup> photons per s) (Howard et al., 1987; Juusola and Hardie, 2001). This performance is remarkable when compared, for example, to amphibian rods, which generate quantum bumps of ~1 pA amplitude with a duration in excess of 1 s and which saturate with photon fluxes of ~500 s<sup>-1</sup> (Baylor et al., 1979). Previous studies have indicated that quantum bumps in *Drosophila* are triggered downstream of PLC, the final amplification being mediated by Ca<sup>2+</sup> influx (Henderson et al., 2000). Bump latency however, is limited by the rate of activation of PLC, since hypomorphic mutations in PLC result in normal size quantum bumps but with prolonged latencies (Scott and Zuker, 1998; Cook et al., 2000). Bump latency is the main constraint on response speed in *Drosophila* (Juusola and Hardie, 2001); our results suggest that short latencies are achieved by exceptionally high PIP<sub>2</sub>-hydrolysis rates consuming much of the immediately available PIP<sub>2</sub> within one microvillus. The rate of hydrolysis required is so great however, that without Ca<sup>2+</sup>-dependent feedback, not only the immediately available PIP<sub>2</sub> but also the entire PI reservoir of up to 4 microvilli is consumed within 1 s. The microvillar design of the rhabdomeric photoreceptor ensures that Ca<sup>2+</sup> influx elicited by a single photon raises Ca<sup>2+</sup> to extremely high levels during the time course of the quantum bump

(Postma et al., 1999). We recently showed that this  $\text{Ca}^{2+}$  influx is required for the facilitation responsible for large quantum bumps (Henderson et al., 2000); the present study suggests that it is also essential to rapidly quench PLC activity and accelerate  $\text{PIP}_2$  recycling, so as to avoid an otherwise debilitating depletion of substrate. For a short period (50–100 ms), the microvillus then becomes refractory (Hochstrate and Hamdorf, 1990) until  $\text{Ca}^{2+}$  is extruded by the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger. We suggest that this allows time for  $\text{PIP}_2$  to be rapidly replenished and also for molecular inactivation of rhodopsin and the G-protein-PLC complex (Cook et al., 2000). When  $\text{Ca}^{2+}$  feedback fails, as in the *trp* mutant, this strategy leaves the PI and  $\text{PIP}_2$  pool in the microvilli vulnerable to total depletion resulting in response decay and inactivation.

#### Experimental Procedures

##### Flies

Flies (*Drosophila melanogaster*) were raised in the dark at 25°C. The wild-type strain was *w* Oregon; mutants included: *trp*<sup>P343</sup> (null; Niemeyer et al., 1996); *rdgB*<sup>K5222</sup> (severe hypomorphic; Milligan et al., 1997); and *cds*<sup>1</sup> (null, Wu et al., 1995). Flies expressing a UAS-*Kir2.1* transgene tagged with an inframe enhanced green fluorescent protein (*p[UAS-EGFP-Kir2.1]*) were generated as described elsewhere (Baines et al., 2001). Two independent transformants (*Kir1*, on the second, and *Kir7*, on the third chromosome) were used with no obvious differences. To induce expression in the photoreceptors (*p[UAS-EGFP-Kir2.1]*) flies were crossed to flies expressing a 3kB fragment containing the promoter for R1–6 rhodopsin (*Rh1*) fused to *GAL4* (*p[Rh1GAL4]*) (again, 2 independent transformants on the 2<sup>nd</sup> and 3<sup>rd</sup> chromosomes were used; provided by J. Treisman and F. Pichaud) and experiments performed on F1 adults. Expression of *EGFP-Kir2.1* was confirmed by GFP fluorescence on dissociated ommatidia, or in the intact animal by using optical neutralization of the cornea (Franceschini and Kirschfeld, 1971) whereby an immersion objective is lowered onto the cornea of an immobilized fly and focused through the facet lenses at the levels of the rhabdomere tips (Figure 2A).

##### Whole-Cell Recordings

Dissociated ommatidia were prepared as previously described from recently enclosed adult flies (Hardie, 1991; Hardie, 1996) and transferred to the bottom of a recording chamber on an inverted Nikon Diaphot microscope. Unless otherwise stated, the bath was composed of (in mM): 120 NaCl, 5 KCl, 10 TES, 4  $\text{MgCl}_2$ , 1.5  $\text{CaCl}_2$ , 25 proline, and 5 alanine.  $\text{Ca}^{2+}$ -free bath was identical except  $\text{CaCl}_2$  was omitted and 250  $\mu\text{M}$  EGTA added (free  $[\text{Ca}^{2+}] < 100$  nM measured fluorimetrically with 5  $\mu\text{M}$  INDO-1). In some experiments, the monovalent cation composition was 110 NaCl, 10 KCl, and 4 CsCl to reduce the amplitude and voltage dependence of the IRK currents. For experiments in  $\text{Ca}^{2+}$ -free bath or  $\text{Na}^+$ -free Ringer, cells were plated in normal bath and perfused shortly before the experiment from a broad tipped (~5–10  $\mu\text{m}$ ) picospritzer. Intracellular solution was (in mM): 140 K gluconate, 10 TES, 4 Mg ATP, 2  $\text{MgCl}_2$ , 1 NAD, and 0.4 Na GTP. pH of all solutions was 7.15. Whole-cell voltage clamp recordings were made using electrodes of resistance ~10 M $\Omega$ ; series resistance values were generally below 25 M $\Omega$  and were routinely compensated to 80%. Data were collected and analyzed using an Axopatch 1-D amplifier and pCLAMP 6 or 8 software (Axon Instruments, Foster City, California). Cells were stimulated via a green LED, with maximum effective intensity of  $\sim 2 \times 10^5$  photons  $\text{s}^{-1}$  per photoreceptor. Relative intensities were calibrated using a photomultiplier and converted to absolute intensities in terms of effectively absorbed photons by counting quantum bumps at low intensities (e.g., Henderson et al., 2000).

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